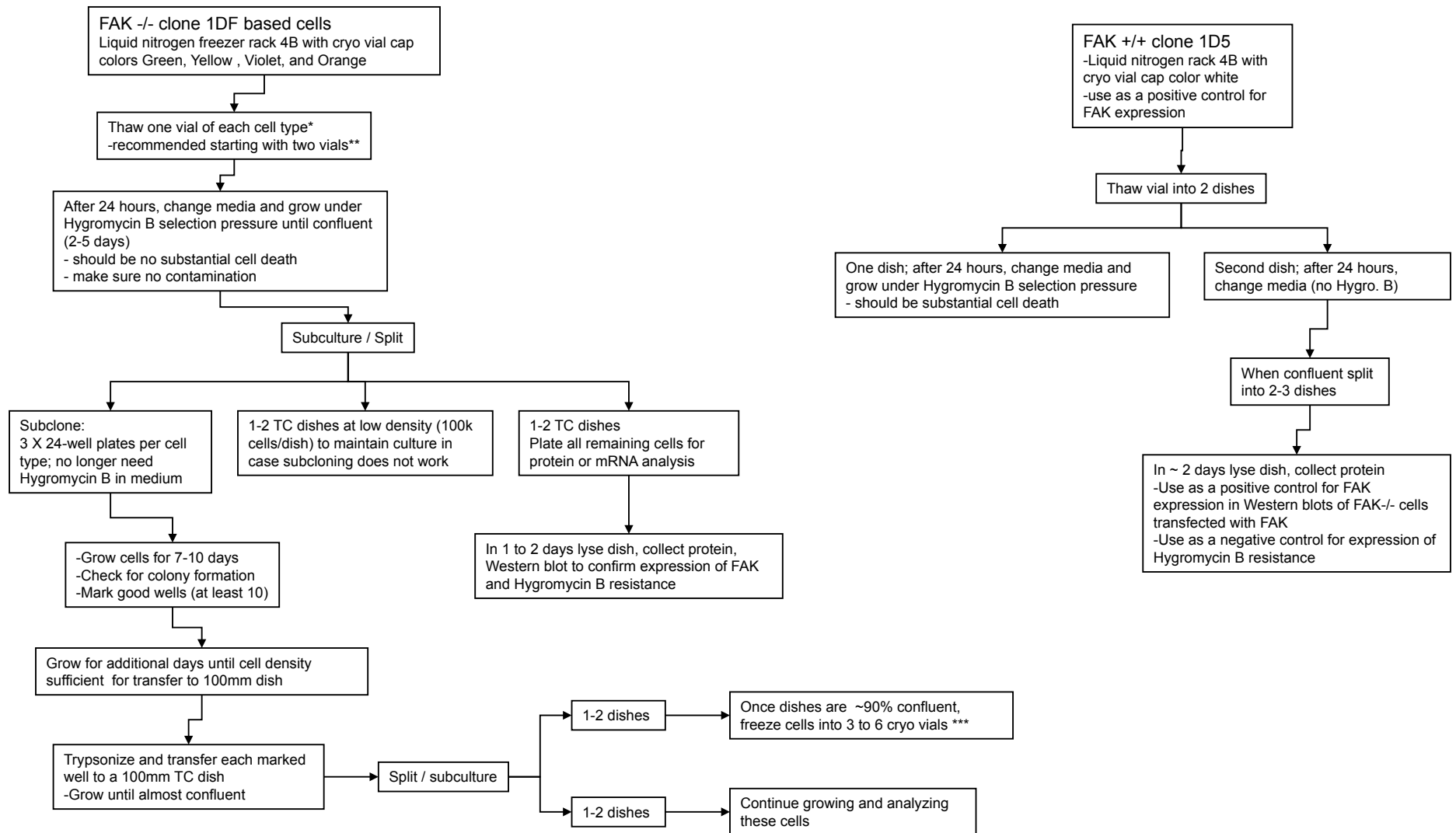


Suggested Outline for Developing Subclones of Transfected FAK ^{-/-} cells



* It is recommended that subcloning of only two types of cells is performed at a time since the final stages of the process will result in 30 + tissue culture dishes per cell type. There are four types of cells developed from FAK ^{-/-} clone 1Df. The FAK ^{-/-} 1Df cells transfected with the empty pcDNA 3.1 Hygro + vector only ("vector control" cells) have **green** cryovial caps and express the Hygromycin B resistance gene only. The FAK ^{-/-} 1Df cells transfected with pcDNA 3.1 Hygro + wtFAK have **violet** cryo vial caps and express the Hygromycin B resistance and wild type murine FAK. In a similar way the cryovials with yellow caps express FAK Y397F mutant (tyrosine 397 change to phenylalanine) and Hygro B resistance, and the cryo vials with **orange** caps express FAK Y925F and Hygro B resistance.

** It is also recommended that you perform this process with the FAK ^{-/-} 1Df cells that are expressing vector only (green) and wtFAK (violet). The wtFAK cells should be the easiest to analyze for the expression of FAK and the vector control cells can be used as a negative control for FAK expression.

*** Several vials of each subcloned cell type should be frozen at the lowest passage number possible. These cells should only be used in the future for growing and freezing more cells (sort of like a farmer's "seed" stock of a certain plant).

Suggested Outline for Developing Subclones of Transfected FAK ^{-/-} cells

FAK ^{-/-} clone 1E6 based cells
Liquid nitrogen freezer rack 4B with cryo vial cap
colors Pink, Gray, Blue, and Tan

Thaw vial
-Note these cells are at a lower
density than FAK ^{-/-} clone 1Df
based cells

After 24 hours, change media and grow under
Hygromycin B selection pressure for a
minimum of 15 days
- These cells need the additional time under
selection pressure to ensure they are stably
transfected
- make sure no contamination

Subculture / Split

Follow procedure given for FAK ^{-/-} 1Df
cells on previous slide for subcloning etc.

Simple illustration of subcloning process

